electrophilic center at the binding site of the enzyme. An alternative to these explanations is that the observation of the lower Michaelis constants for the halogensubstituted dipeptides may depend in some way on the electron-withdrawing inductive effect of the halogen atoms on the aromatic ring of the tyrosine nucleus. In this regard the powerful influence of the bromo and iodo substituents is evidenced by the pK_a' OH values measured for the substituted phenolic groups as shown in Table I. The pK_a' OH values for the hydroxyl groups of the tyrosine residues in N-acetylphenylalanyl-L-3,5-dibromotyrosine and N-acetylphenylalanyl-L-3,5diiodotyrosine are much lower than that in the unsubstituted dipeptide.

Another point which requires explanation is that in a recent report of work on the pepsin-catalyzed hydrolysis of N-acetyl-L-phenylalanyl-L-diiodotyrosine, inhibition by the product, N-acetyl-L-phenylalanine, was not found⁶ despite the observation of competitive inhibition by the same product in studies on the hydrolysis of N-acetyl-L-phenylalanyl-L-tyrosine³ (see Table I). In our investigation of the hydrolysis of N-acetyl-L-phenylalanyl-L-3,5-dibromotyrosine, we found inhibition by N-acetyl-L-phenylalanine although the value of $K_{\rm I}$ which we measured cannot be compared directly to that observed in the hydrolysis of N-acetyl-L-phenylalanyl-L-tyrosine since Silver, et al., 3 did not report a value for $K_{\rm I}$.

In summary, our results indicate that the hydrolytic behavior of the dibromo dipeptide lies in between that observed for the diiodo dipeptide and that of the sub-



Figure 1. Lineweaver-Burk plot for the hydrolysis of N-acetyl-Lphenylalanyl-L-3,5-dibromotyrosine by $7 \times 10^{-7} M$ pepsin at pH 2.0.

stituted dipeptide. In order to further elaborate on the causes of trends in both $K_{\rm m}$ and $k_{\rm cat}$, we are systematically varying the polarizability and size of substituents. For these reasons the kinetics of the pepsin-catalyzed hydrolysis of N-acetyl-L-phenylalanyl-L-3,5-dinitrotyrosine and related dipeptides are now under study.

Communications to the Editor

A Total Synthesis of a Natural Prostaglandin

Sir:

Prostaglandins, a family of C₂₀ prostanoic acids of the same carbon skeleton¹ first isolated by von Euler² and Goldblatt³ as crude extracts from sheep seminal vesicular glands or human seminal plasma, are now known to be physiologically important lipids^{1,4-6} widely distributed in nature.^{1,7} In an elegant series of chemical, spectral, and crystallographic studies Bergstrom and co-workers first isolated PGE₁ (prostaglandin E_1 , (15R)-9-oxo-11 α , 15-dihydroxyprost-13trans-enoic acid,1c in pure form8 and established its

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structure (I).^{1a,b,9,10} We now report the synthesis (see Scheme I) of the racemic ethyl ester of (15R)-9-oxo- 11α , 15-dihydroxyprostanoic acid (dihydro-PGE₁, XIVa), a biologically potent^{11,12} naturally occurring¹³ metabolite of PGE_1 .

Formylation (sodium hydride-ethyl formate) of 3-ethoxy-2-cyclopentenone (II)¹⁴ afforded nearly quantitative yields of the sodium salt of the 5-hydroxymethylene derivative III. Heterogeneous reaction of III in chloroform with ethyl bromoacetate and triphenylphosphine gave IV¹⁵ in 70% yield, mp 64-65°, $\lambda_{\text{max}}^{\text{EtoH}}$ 251 (ϵ 14,500) and 260 m μ (ϵ 14,300), prepared also, after neutralization of III (although less conveniently), with carbethoxymethylenetriphenylphosphorane.

Acid hydrolysis of IV yielded the enol Va¹⁵ [mp 155-160°, $\lambda_{max}^{Et\delta H}$ 275.5 m μ (ϵ 17,900)] which, after treatment

(9) S. Bergström, R. Ryhage, B. Samuelsson, and J. Sjöval, ibid., 16, 501 (1962).

(10) (a) S. Abrahamson, S. Bergström, and B. Samuelsson, Proc. Chem. Soc., 332 (1962); (b) S. Abrahamson, Acta Cryst., 16, 409 (1963). (11) E. Anggård, Acta Physiol. Scand., in press.

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(13) E. Änggård and B. Samuelsson, J. Biol. Chem., 239, 4097 (1964). (14) H. O. House and G. H. Rasmusson, J. Org. Chem., 28, 27 (1963).

(15) Analysis within journal requirements and compatible nmr spectra.

⁽¹⁾ For leading references and nomenclature see: (a) S. Bergström and B. Samuelsson, Ann. Rev. Biochem., 34, 101 (1965); (b) B. Samuelsson, Angew. Chem. Intern. Ed. Engl., 4, 410 (1965); (c) B. Samuelsson, J. Biol. Chem., 238, 3229 (1963); (d) E. W. Horton, Experientia, 21, 113 (1965).

Scheme I



^a In compounds VI and IX, the side chain is unsaturated (dotted bonds). VII, VIII, X, and XI have saturated side chains.

with diazoethane, gave Vb¹⁵ [mp 48–50°; λ_{max}^{EtoH} 206 (ϵ 12,400), 211 (ϵ 11,400), and 269.5 m μ (ϵ 16,600)]. Ultraviolet maxima of IV and Vb confirm the structures assigned and hence the structure of III.

In a key extension of the Wittig reaction, addition of III to a solution of ethyl 6-bromosorbate¹⁶ and triphenylphosphine yielded VI¹⁵ [mp 85–88°, λ_{max}^{EtOH} 216 (ϵ 10,750), 252 (ϵ 8650), and 327.5 m μ (ϵ 38,950)] in 35–50% yield. Catalytic hydrogenation selectively reduced the side chain to give VII.¹⁵ Again, formylation (sodium hydride–ethyl formate) produced the sodium salt of the hydroxymethylene derivative VIII which reacted with *n*-hexanoylmethylenetriphenylphosphonium chloride¹⁷ in chloroform to give IX¹⁵ as a mixture of geometric isomers (48% yield). Hydrogenation of the exocyclic bond (5% palladium–charcoal; trace of triethylamine) proceeded in 85% yield to give X.^{15,18} Acid-catalyzed solvolysis of X in benzyl alcohol gave

Acid-catalyzed solvolysis of X in benzyl alcohol gave XI¹⁵ and the isomeric enol ether. Without separation, XI was reduced with lithium tri-*t*-butoxyaluminum hydride to the corresponding 15-hydroxy compound XII^{15,19} in 85% yield. Catalytic hydrogenolysis of this mixture (5% palladium–charcoal) gave the β -diketone XIII which was subjected to prolonged catalytic hydrogenation (30% rhodium–charcoal) to produce a complex mixture. A major component of this mixture, XIVa,¹⁵ obtained in 11% yield as a fraction homo-

(16) H. E. Ungnade and T. R. Hopkins, J. Am. Chem. Soc., 73, 3091 (1951).

(17) S. Trippett and D. M. Walker, J. Chem. Soc., 1266 (1961).

(18) Isomerization of the Δ^{12} to the less-hindered Δ^{13} bond is believed to occur during reduction of IX to X in the presence of base. Although X contains two asymmetric centers, it is largely one racemate with *trans* substituents, homogeneous by tlc and with a well-defined nmr spectrum. Mass spectral and labeling studies confirm this stereochemical preference.

(19) This compound is undoubtedly a mixture of diastereoisomers as little stereochemical control appears likely here.

geneous on tlc, exhibited a polarity identical with that of authentic dihydro-PGE₁ ethyl ester (XIVc).

Although both materials were oils, XIVa was shown both to have the correct gross structure and to include material of proper stereochemistry in the following manner.²⁰

Both compounds exhibited the same mobility on tlc, possessed identical infrared spectra, underwent alkaline isomerization^{1b,21} to products (λ_{max} near 240 m μ) with the same mobility on tlc, had common nmr maxima with no extraneous peaks, showed the same parent ion at m/e 384 and prominent fragments at m/e 366, 348, 295, 210, and 192 in the mass spectrum,^{22,23} exhibited muscle-stimulating and vasodepressor activity in the guinea pig ileum and pentolinium-blocked dog as-says^{24,25} of the same order of magnitude,¹² and were dehydrogenated enzymatically by the 15-dehydrogenase from pig lung.^{11,26,27}

Final confirmation of structure and stereochemistry of a component of the final mixture was obtained by radioisotope dilution. Authentic crystalline methyl

(21) S. Bergström, R. Ryhage, B. Samuelsson, and J. Sjövall, J. Biol. Chem., 238, 3555 (1963).

(22) Especially significant were peaks at m/e 210 and 192 formed by loss of the acid side chain through McClafferty rearrangement: F. W. McClafferty, *Anal. Chem.*, 31, 82, 1959.

(23) The similarity of the mass spectra is considered to support the *trans* side chain configuration since *cis*- and *trans*-dialkylcyclopentanes have been reported to give different mass spectra: J. Momigny and P. Natalis, *Bull. Soc. Chim. Belges*, **66**, 26 (1957).

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(26) S. Bergström and B. Samuelsson, private communication.

(27) E. Änggård and B. Samuelsson, Arkiv Kemi, 25, 293 (1966).

⁽²⁰⁾ As this material occasionally decomposed on standing, the final product was freshly purified before most of the following comparisons were made.

(15R)-9 β ,11 α ,15-trihydroxyprostanoate (dihydro-PGF₁₈) methyl ester, XVb),¹⁵ mp 62-63°, was prepared from PGE_1 (I) by catalytic hydrogenation^{9, 28} and reduction with sodium borohydride. 29

The synthetic racemic ketoester, XIVa, was reduced with sodium borotritide, hydrolyzed, and reesterified with diazomethane, yielding a mixture of 9-tritiated-9hydroxy epimers. A sample (XVa) enriched in the 9β -hydroxy isomer by chromatography on silica gel was combined with optically active, crystalline XVb. Two crystallizations afforded material of constant melting point (62-63°) and specific activity (0.249 μ curie/mg) unchanged on two further crystallizations (0.238 and 0.236 µcurie/mg). Isotope dilution indicated that XIVa contained at least 22% of product of the same optical configuration at all asymmetric centers as XIVc. 30,31

Acknowledgment. The authors wish to express their appreciation to their associates in these laboratories, especially Marvin Grostic for his contributions in mass spectroscopy, Byron Johnson for preparation of intermediates, and George Cartland and James Weeks for biological studies.

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(29) S. Bergström, L. Krabisch, B. Samuelsson, and J. Sjövall, ibid., 16, 969 (1962).

(30) Stereochemical randomness would produce only 6% of the natural isomer in the introduction of four asymmetric centers.

(31) NOTE ADDED IN PROOF. D. H. Nugteren and D. A. vanDorp, Unilever Laboratories, and B. Samuelsson and S. Bergstrom, Karolinska Institute, announced independently at the 2nd Nobel Symposium on The Prostaglandins, June 6, 1966, that reexamination of the original data indicates that the absolute configuration of the prostaglandins should now be represented by the mirror image of that used in this paper and in earlier publications.

> P. F. Beal, III, J. C. Babcock, F. H. Lincoln Research Laboratories, The Upjohn Company Kalamazoo, Michigan Received May 27, 1966

Ground- and Transition-State Free Energy Relationships in σ and π Routes to the Nonclassical 7-Norbornenyl Cation¹

Sir:

One of the most interesting of the nonclassical carbonium ions is the intermediate II in solvolysis of anti-7-norbornenyl derivatives² such as I-OTs. In this communication we report on kinetic and thermodynamic product control and ground- and transition-state free energy relationships in σ and π routes³ to cation II in methanolysis.

A tricyclic derivative suitable for the σ route to cation II, namely, III-OCH₃, was obtained very recently in an elegant investigation by Tanida, Tsuji, and Irie,4ª who observed considerable tricyclic product IV-OCH₃ in methanolysis of 7-norbornadienyl chloride in the presence of substantial concentrations of sodium methoxide. Hydrogenation of IV-OCH₃ led to III- OCH_3 , an 85% pure sample of which was isolated. Tanida and co-workers^{4a} also reported tricyclic III-

Table I. Kinetic Product Control in Methanolysis of I-OTs^a at 25.0°

[NaOCH ₃], <i>M</i>	% I -OC H₃	% III–OCH3
	100	
3.5×10^{-8}	99.7	0.3ª
0.20	93.6	6.4
0.88	79.7	20.3
1.93	65.3	34.7
3.98	48.5	51.5

^a Ca. 0.01 or 0.02 M I-OTs. ^b Solution allowed to become acidic. ° 0.0452 M NaOAc-0.0394 M AcOH buffer; (MeO-) varies from 7.5 \times 10⁻⁸ M to 3.8 \times 10⁻⁸ M. ^d This material disappears in acidic 80% acetone with the same half-life as for III-OCH₃.

OCH₃ from methanolysis of I-OTs with alkali, but in only 1 % yield.

We noted and remarked previously^{2a} on the fact that the $C_7: C_2$ reactivity ratio in cation II is quite dependent on the nature of the nucleophile when it is varied widely. Thus, tricyclic alcohol III-OH was not observed^{2a} from neutral hydrolysis of I-OTs, but considerable tricyclic hydrocarbon III-H is obtained from borohydride trapping,² and some tricyclic nitrile III-CN is formed in the reaction of I-Cl with NaCN in aqueous alcohol.4b We have now observed striking changes in product composition in methanolysis of I-OTs as methoxide concentration is varied. In methanol at 25.0°, I-OTs undergoes first-order methanolysis with a rate constant of $1.31 \times 10^{-3} \text{ sec}^{-1}$ to form I-OCH₃ quantitatively. No tricyclic III-OCH₃ is observed, but even if it were formed, it would not survive the acidic conditions since it is very easily isomerized to I-OCH₃ in the presence of acid. As summarized in Table I, methanolysis of I-OTs with increasing concentrations of NaOCH₃ gives increasing proportions of tricyclic III-OCH₃ in the product. Thus, with 4 M NaOCH₃, the observed ether contains 51.5% tricyclic III-OCH₃. The data make it evident that sodium methoxide does not serve merely to preserve the tricyclic III-OCH₃, but instead is largely responsible for its formation.



The kinetic control product from neutral methanolysis of I-OTs was determined from solvolysis in methanol containing an AcOH-NaOAc buffer. Control experiments showed the tricyclic III-OCH₃ to be fully stable under these conditions. The product ether from buffered methanolysis contained 0.3% of the tricyclic III-OCH₃, as determined by vpc. Examination of the nmr spectrum, using a Varian C-1024 time-averaging computer (CAT), confirmed the presence of III-OCH₃ with its characteristic $C_2 \alpha$ -proton quartet centered at τ 6.35. Thus, kinetic product control in methanolysis of I-OTs involves a C7:C2 reactivity ratio⁵ of ca. 300.

(5) The situation regarding kinetic and thermodynamic control in

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